

# From Robots to Proteins: Randomized Motion Planning for High-Dimensional Problems

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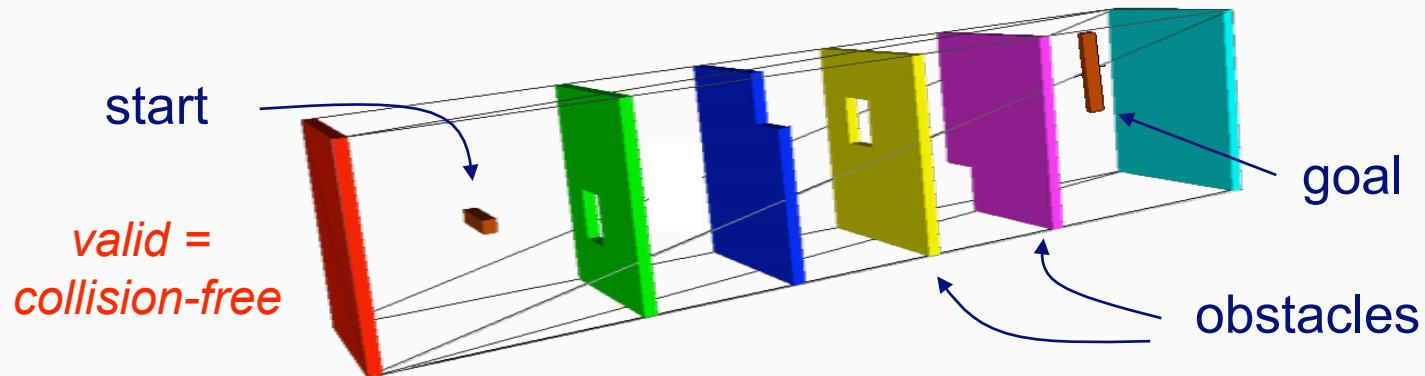
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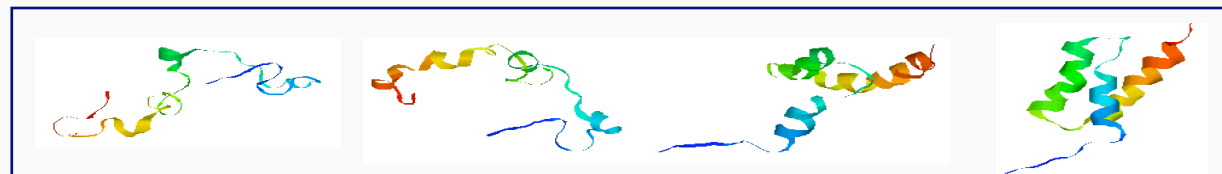
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# What is motion planning?

- Find a **valid path** from a **start** to a **goal** for a movable object

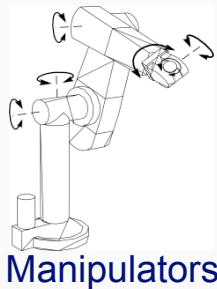


valid =  
low energy



# Motions: Robots, Graphics, Molecules

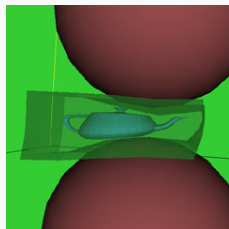
- What do all of these have in common?



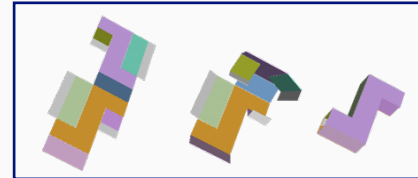
Manipulators



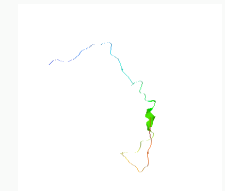
Mobile robots



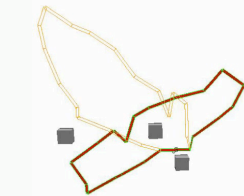
Deformation



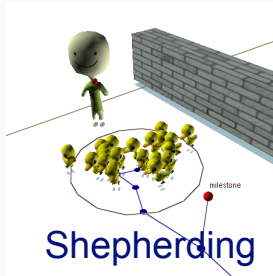
Paper folding



Protein folding



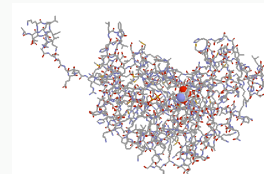
Closed chains



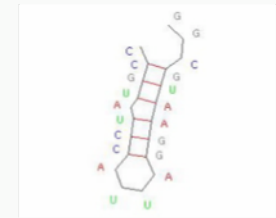
Shepherding



Flocking



Drug docking

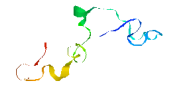


RNA folding

- They are all examples of the motion planning problem
- They can all be solved with the same framework!

# Why Study Folding Pathways?

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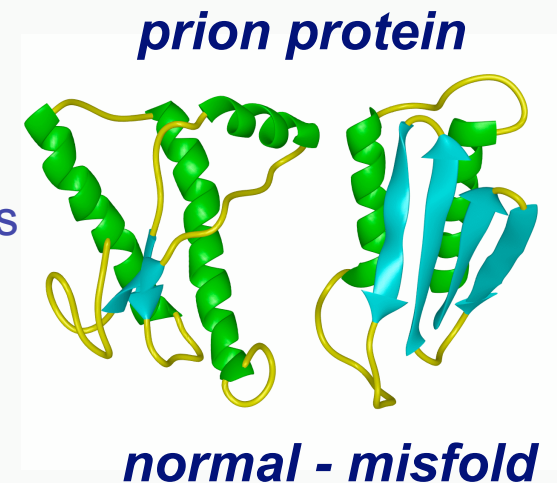


## Importance of Studying Pathways

- Insight into protein interactions & function
  - May lead to better structure prediction algorithms
- Diseases such as Alzheimer's & Mad Cow related to misfolded proteins

## Computational Techniques Critical

- Hard to study experimentally (happens too fast)
- Can study folding for thousands of already solved structures
- Help guide/design future experiments

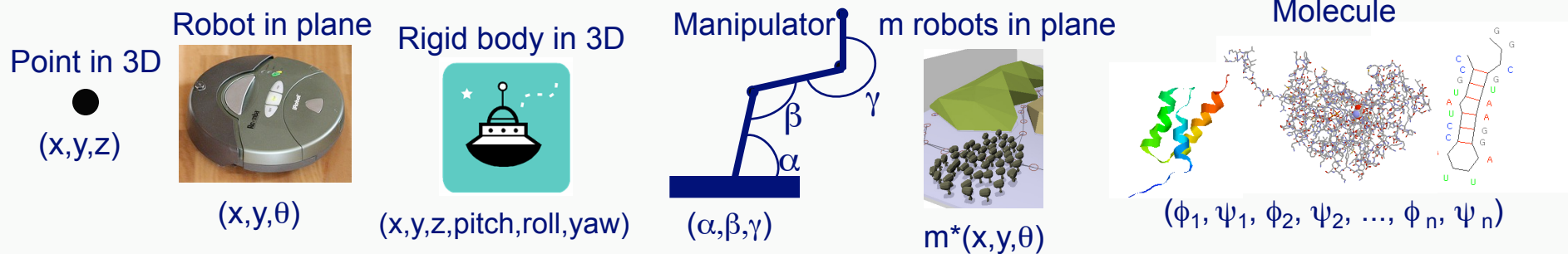


# Motion Planning Framework

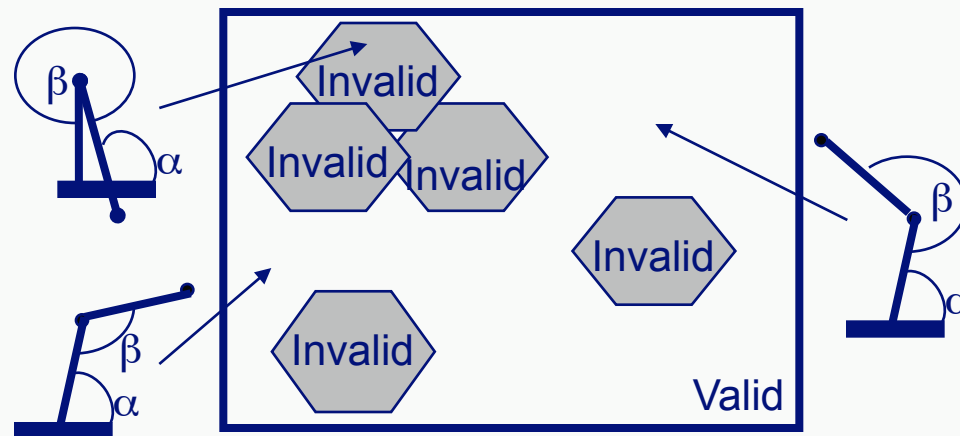
## Robot Abstraction



- How can we develop a single framework to solve all of these different problems?



**Configuration Space (C-space):**  
the set of all object placements



# Motion Planning Framework

## Probabilistic Roadmap Methods (PRMs)



[Kavraki, Svestka, Latombe, Overmars 1996]

- Idea: Build a model (roadmap) that approximates the topology of the space of Configurations

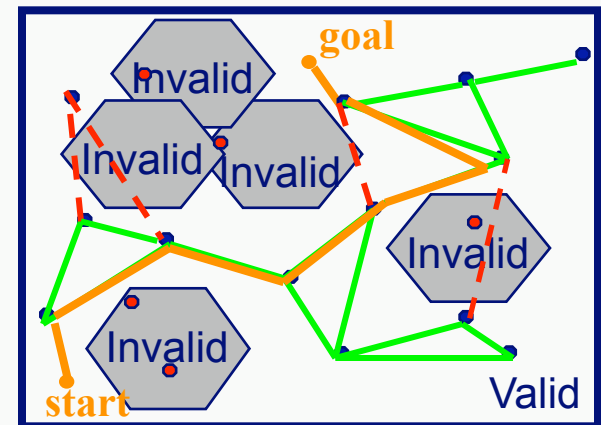
### Roadmap Construction

1. Randomly generate robot samples (nodes)
  - discard nodes that are invalid
2. Connect node pairs to form a **roadmap**
  - simple *local planner*
  - discard paths (edges) that are invalid

### Query processing

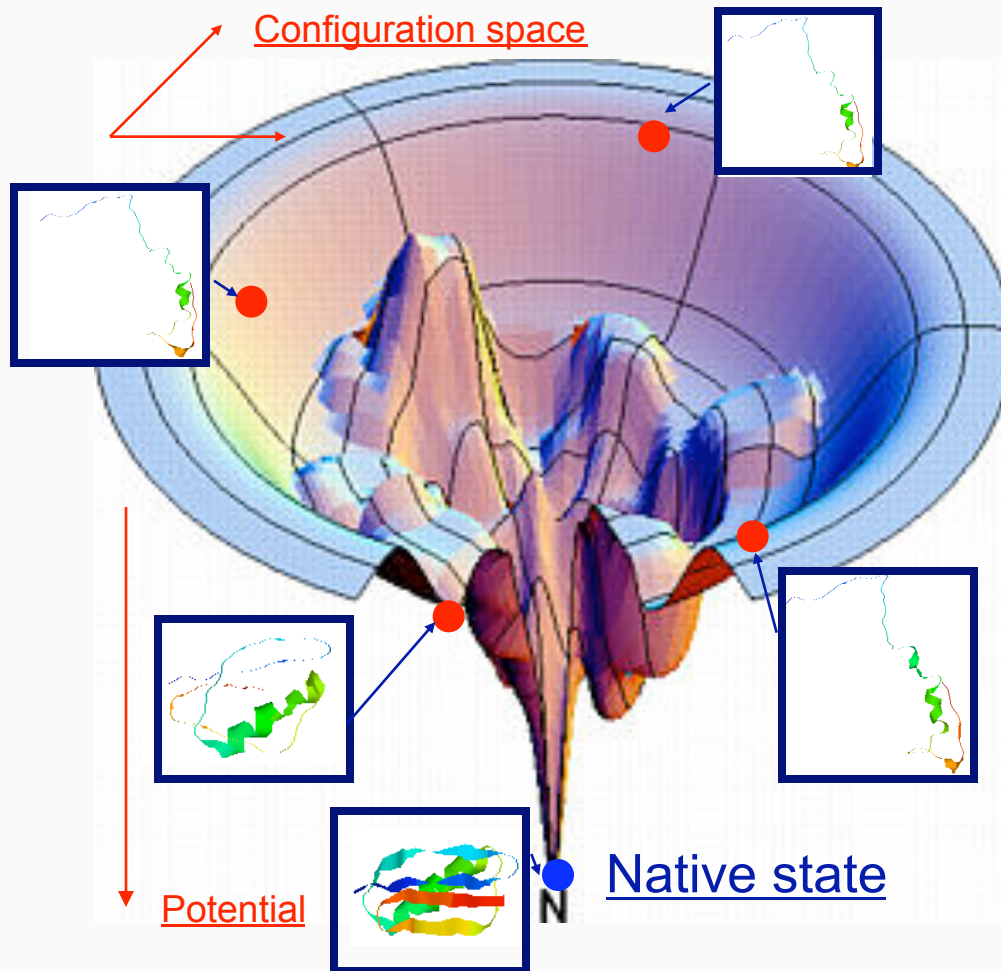
1. Connect *start* and *goal* to roadmap
2. Find path(s) in roadmap between *start* and *goal*

### C-space



***There's something unique about the space***

# The Protein Folding Landscape



## Potential Energy Landscape

- Funnel shape
- Native state is global minimum
- Different proteins  $\Leftrightarrow$   
Different landscapes  $\Leftrightarrow$   
Different folding behaviors

Goal: Build a model (roadmap) of the energy landscape

- Characterize main features
- Extract folding pathways
- Extract folding kinetics

*The energy landscape is huge!*

# Related Work

## Simulating Folding & Kinetics



	Approach	Folding Landscape	# Paths Produced	Path Quality	Compute Time	Folding Kinetics
Trajectory based	Molecular Dynamics [Levitt 83; Haile 92; Daggett, Levitt 93; Duan & Kollman, 98; Shirts & Pande 00, Boczko & Brooks 95]	No	1	Good	Long	Yes
	Monte Carlo Simulation [Covell 92; Kolinski, Skolnick 94]	No	1	Good	Long	Yes
Statistics based	Master Equation Calculation [Cieplak et al. 98, Ozkan et al. 01, 02, Weikl and Dill 03; Weikl et al. 04]	Yes (required)	N/A	N/A	Fast	Yes
	Statistical Models [Muñoz et.al. 98; Alm, Baker 99; Muñoz, Eaton 99; Baker 00; Matysiak, Clementi 04; Das et al.05]	Yes	0	N/A	Fast	Average
Graph based	SRS and $P_{fold}$ [Apaydin et al. 01, Chiang et al. 06]	Yes	Many	Coarse	Fast	Yes
	<b>Our Roadmap-Based</b> [Song, Amato ICRA 01, JCB 01; Amato et al. JCB 02, Thomas, Tang, <b>Tapia</b> , Amato JCB 07, <b>Tapia</b> , Tang, Thomas, Amato Bioinformatics 07, Thomas, <b>Tapia</b> , AmatoTR08-004, <b>Tapia</b> , Thomas, Amato TR08-005; <b>Tapia</b> , Thomas, Amato CIS 09]	Yes	Many	Approx. (tunable)	Fast	Yes

- Other Roadmap-based approaches for studying molecular motions
  - Ligand binding [Singh, Latombe, Brutlag ISMB 99; Bayazit, Song, Amato ICRA 01]
  - RNA Folding [Tang, Kirkpatrick, Thomas, Song, Amato JCB 05; Tang, Thomas, **Tapia**, Amato RECOMB 07; Tang, Thomas, **Tapia**, Giedroc, Amato JMB 08]



# Preliminaries:

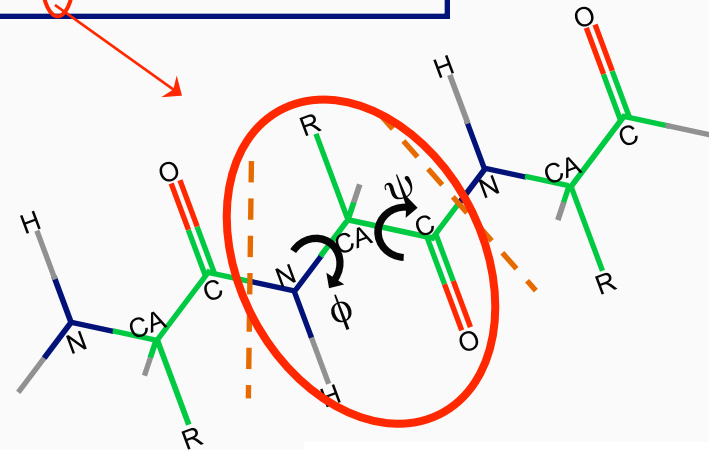
## Protein Structure/Model



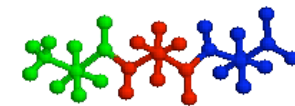
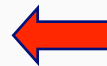
A protein is a sequence of amino acids/residues, each with 2 torsional degrees of freedom



TTCCPSIVARSNFNVCRLPGTPEALCATYTGCIIPGATCPGDYAN



$\{\phi_1, \psi_1, \phi_2, \psi_2, \dots, \phi_n, \psi_n\}$



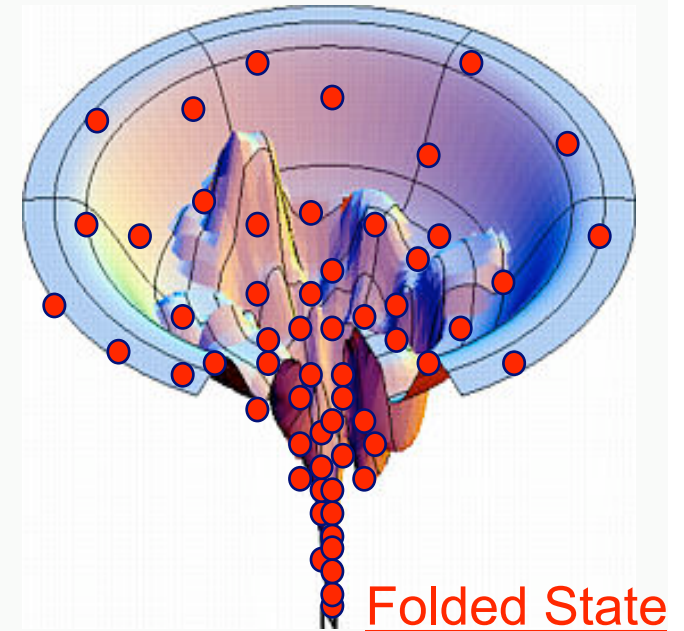
# Protein Folding by Motion Planning

## Node Generation

- Sample using **known target state**
- Criterion for accepting a node:  
Compute potential energy  $E$  of each node and retain it with probability:

$$P(E) = \begin{cases} 1 & \text{if } E < E_{\min} \\ \frac{E_{\max} - E}{E_{\max} - E_{\min}} & \text{if } E_{\min} \leq E \leq E_{\max} \\ 0 & \text{if } E > E_{\max} \end{cases}$$

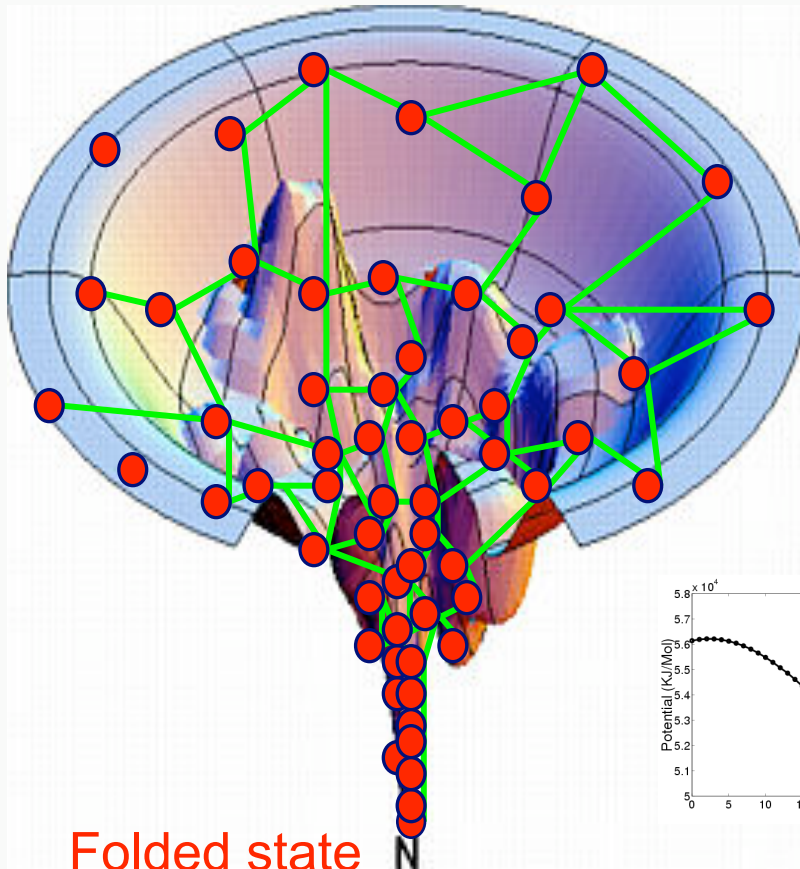
Our coarse energy function is similar to [Levitt 83] and includes van der Waals, hydrogen bonds, and hydrophobic interaction components



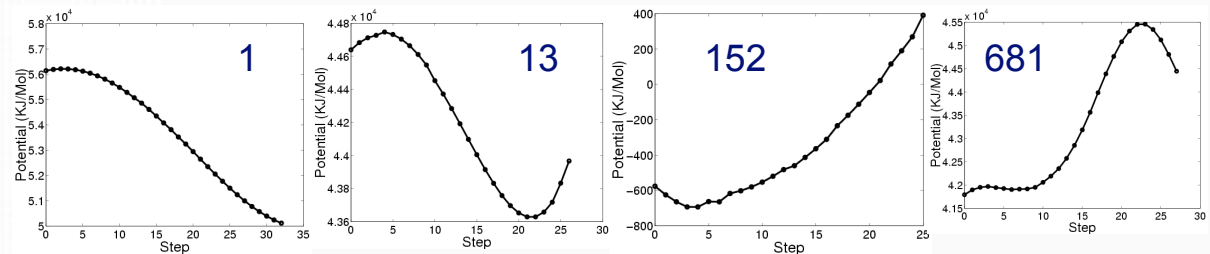
Denser distribution  
around target state  
Biased sampling to  
reduce search  
space

# Protein Folding by Motion Planning

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Node Connection



1. Find  $k$  closest nodes for each roadmap node
  - Conformation space distance metric
  - Euclidean, RMSD, **Rigidity-Based**,...
2. Assign edge weight  $w$  to reflect energetic feasibility



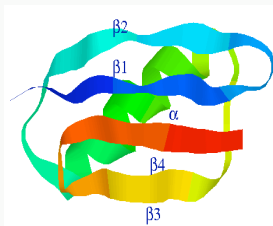
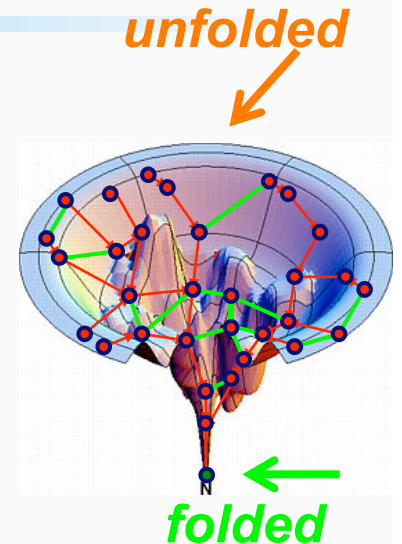
lower weight  $\Leftrightarrow$  more feasible

# Protein Folding

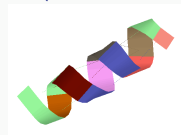
## Path Extraction and Analysis

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- Roadmap contains thousands of folding pathways from **unfolded** to **folded**
  - Extract using Dijkstra's shortest path alg.
  - Analyze pathway's energy profile, secondary structure formation order, etc.
- We group pathways based on their secondary structure formation order



Q: Which forms first?



$\alpha$  helix



$\beta$  sheet

Secondary structure piece is **formed** when it contains most of the native contacts / it is mostly rigid

Do our pathways produce the same orders as seen experimentally?

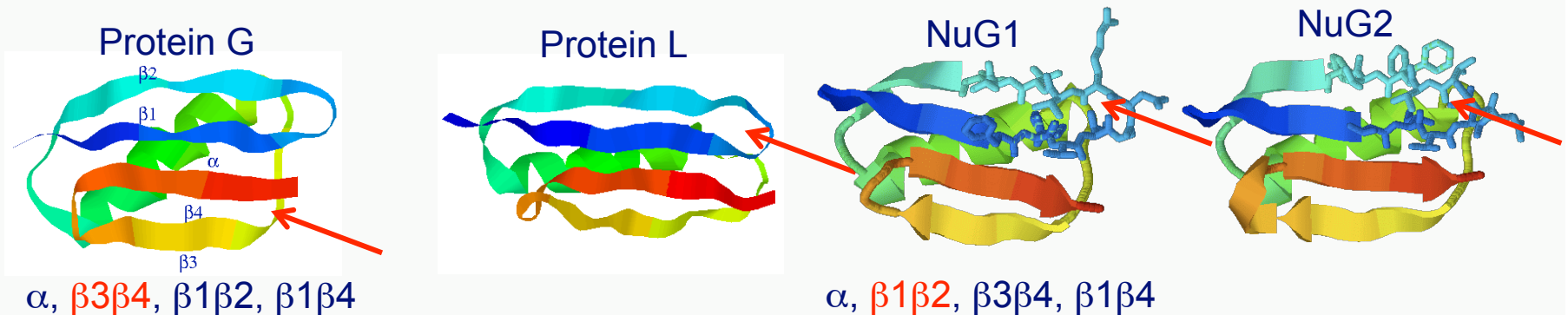
# Protein Folding

## Formation Order of G, L, and Mutants



- Proteins G, L, and two mutants of G (NuG1 and NuG2) have **similar structure** but **fold differently**

[Li, Woodward 99] [Nauli, et al., 01]



Protein	Experimental Order	Roadmap Order	%
G	$[\alpha, \beta 1, \beta 3, \beta 4], \beta 2^1$ $[\alpha, \beta 4], [\beta 1, \beta 2, \beta 3]^2$	$\alpha, \beta 3-4, \beta 1-2$ $\beta 3-4, \alpha, \beta 1-2$	99.4 0.6
L	$[\alpha, \beta 1, \beta 2, \beta 4], \beta 3^1$ $[\alpha, \beta 1], [\beta 2, \beta 3, \beta 4]^2$	$\beta 1-2, \alpha, \beta 3-4$	100.0
NuG1	$\beta 1-2, \beta 3-4^3$	$\alpha, \beta 1-2, \beta 3-4$	97.6
NuG2	$\beta 1-2, \beta 3-4^3$	$\alpha, \beta 1-2, \beta 3-4$	96.6

Folding behavior for all four proteins predicted [Thomas, Tang, Tapia, Amato JCB 07]

Folding rates for G, NuG1, NuG2 are drastically different [Nauli, et al., 01]

<sup>1</sup> Hydrogen out-exchange experiments [Li, Woodward 99]

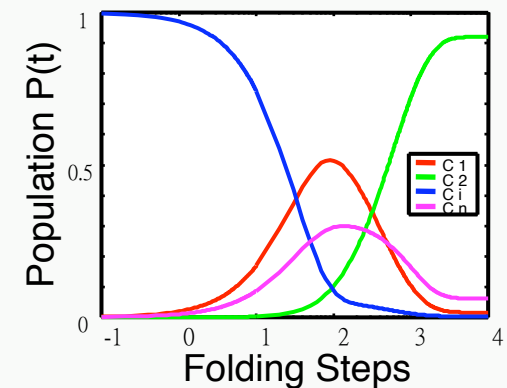
<sup>2</sup> Pulsed labeling/competition experiments [Li, Woodward 99]

<sup>3</sup> F-value analysis [Nauli, et al., 01]

# Protein Folding Kinetics

**Kinetics is the study of reaction rates**

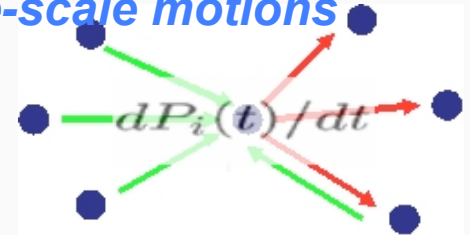
- Folding rates – Faster vs. Slower
- Population kinetics – Change in Conformers
- Validation with Other Experimental Techniques
  - Tryptophan Fluorescence
  - Circular Dichroism
  - H/D Exchange



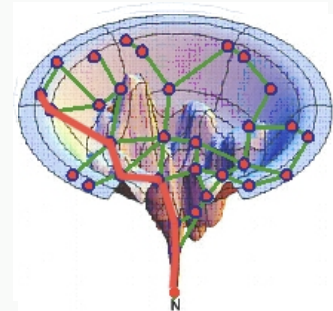
# Map-Based Analysis Techniques

→ *Uses local transition probabilities to identify likely large-scale motions*

Technique 1: Map-Based Master Equation  
Calculation (MME)



Technique 2: Map-Based Monte Carlo (MMC)

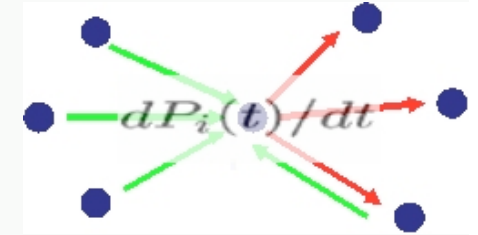


These techniques provide results that  
can be validated against lab experiment!



# Map-Based Technique 1

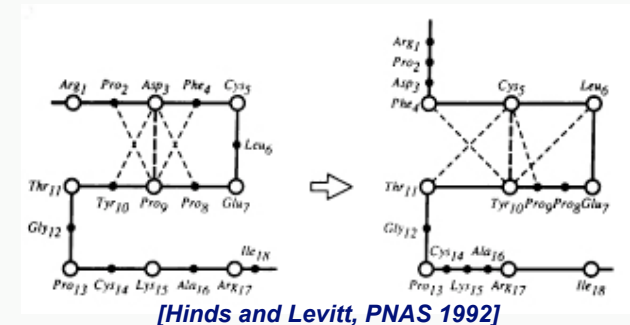
## Map-Based Master Equation (MME)



- **Master Equation (ME)** is a differential equation describing the probability of a process to be in a given state

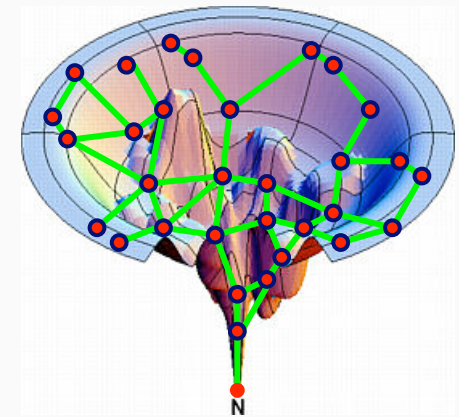
- Challenge:

- Usually applied to a **detailed model** of the energy landscape (lattice, etc.)
- Thus, limited to small proteins



- Our solution:

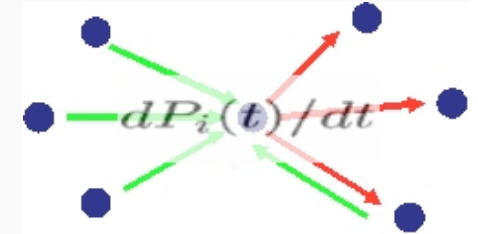
- Apply to our **roadmap** (approximate landscape model) instead
- Roadmap gives model (conformations and transitions) for master equation



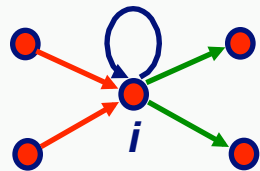


# Map-Based Technique 1

## Map-Based Master Equation (MME)



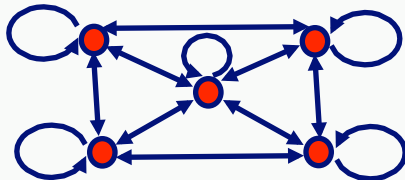
- For conformation  $i$ , its **population over time** can be described by:



$$dP_i(t)/dt = \sum_{i \neq j}^n (\underbrace{k_{ji} P_j(t)} - \underbrace{k_{ij} P_i(t)})$$

$k_{ij}$  is a transition probability calculated from edge  $ij$  in our roadmap

- The master equation describes the population kinetics of all conformations



$$d\mathbf{p}(t)/dt = M\mathbf{p}(t)$$

$$\begin{cases} M_{ij} = \underbrace{k_{ji}} & i \neq j \\ M_{ii} = -\underbrace{\sum_{i \neq j} k_{ij}} \end{cases}$$

- The solution encodes **folding rates** (eigenvalues) and important **conformation distributions** (eigenvectors)

$$P_i(t) = \sum_k \sum_j N_{ik} e^{\lambda_k t} N_{kj}^{-1} P_j(0)$$

$N_{i0}$  = Boltzmann equilibrium distribution

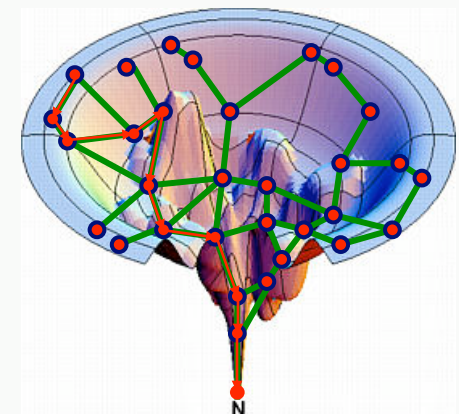
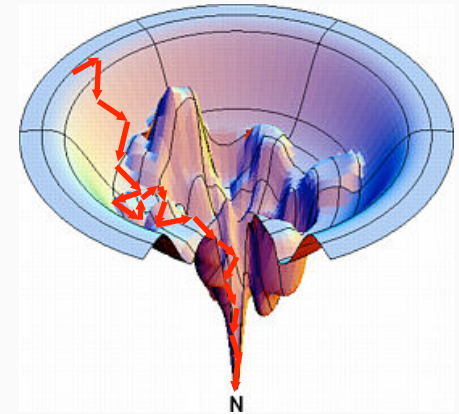
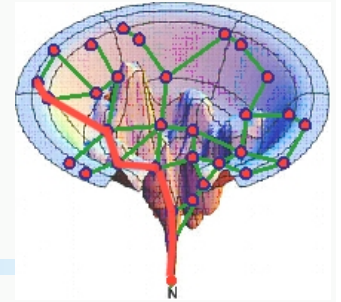
$\lambda_1$  = folding rate (for 2-state folders)

# Map-Based Technique 2



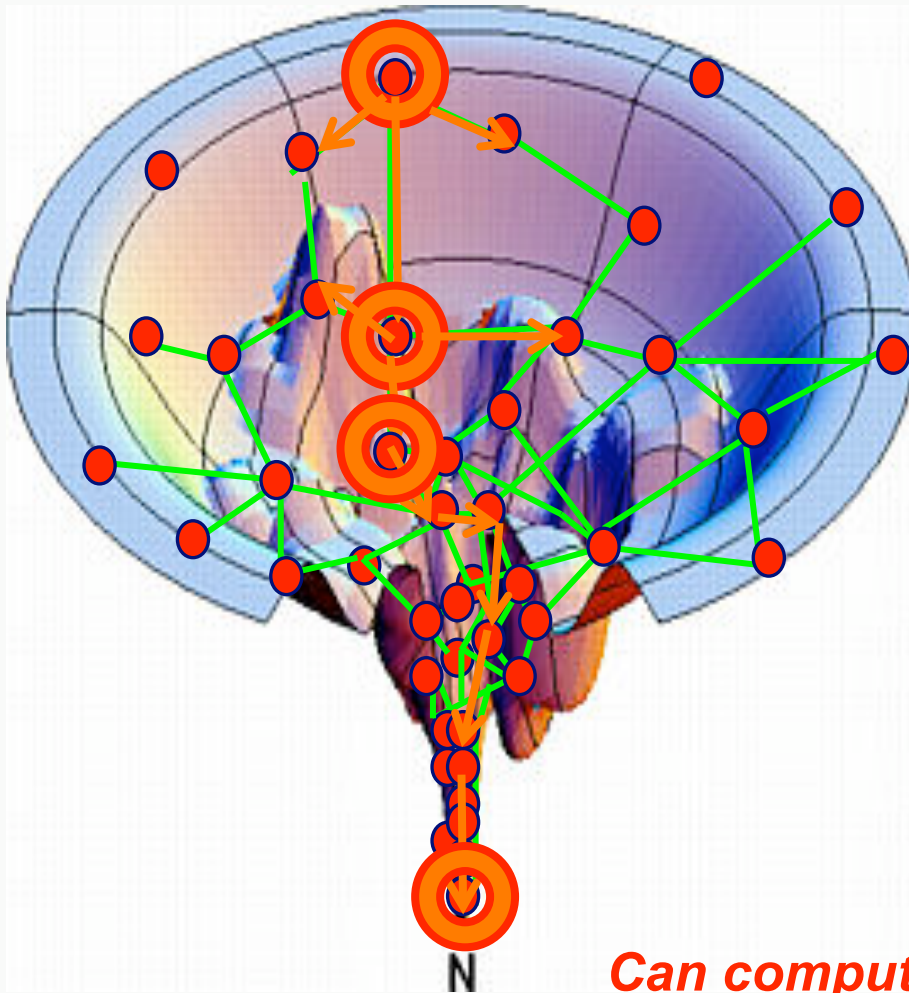
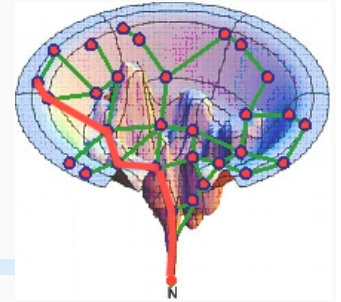
## Map-Based Monte Carlo (MMC)

- **Monte Carlo (MC) simulation** is a random walk on the energy landscape
- Challenge: [Covell, 1992; Kolinski and Skolnick, 1994]
  - At every timestep, MC computes the **complete** local landscape
  - Limited to small proteins
- Our solution:
  - Apply to our **roadmap** (approximate landscape model) instead
  - Calculate structure formation from MMC paths



$P_{ij}$  proportional  
to  $1/w_{ij}$

# MMC Algorithm



- Start at random unfolded state, current node
- Repeat until maximum number of steps
  - Identify adjacent nodes (neighbors) of current node in the map
  - Calculate the transition probabilities from the edge weight
  - Move to a neighbor probabilistically

***Can compute population kinetics and structural features of each conformation in each timestep***

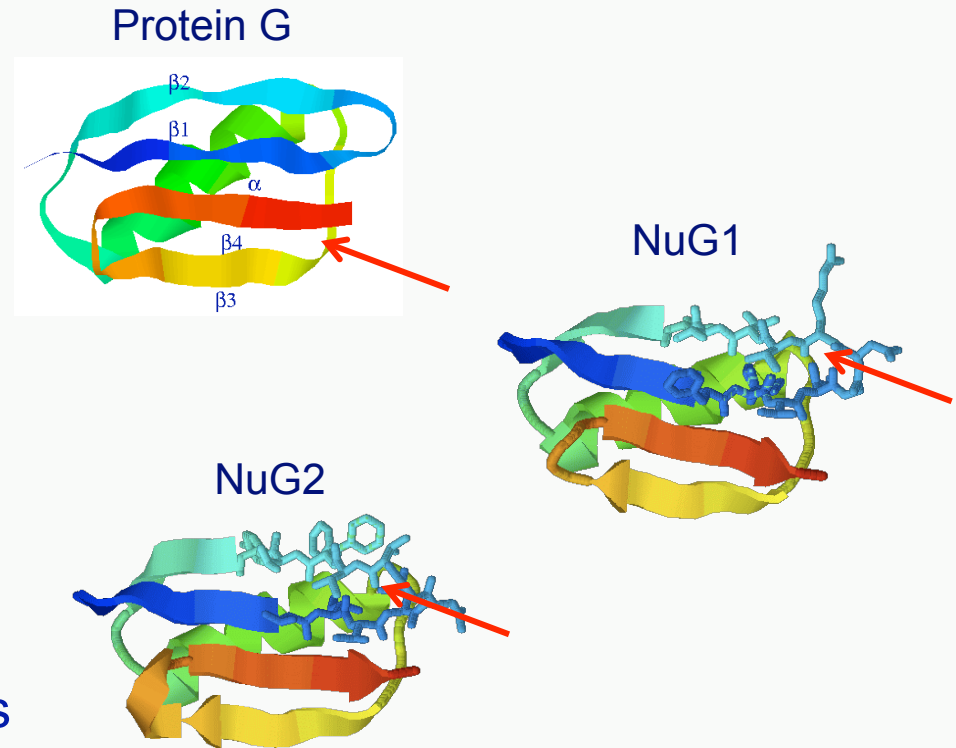
# Kinetic Case Study

## Protein G, NuG1, and NuG2



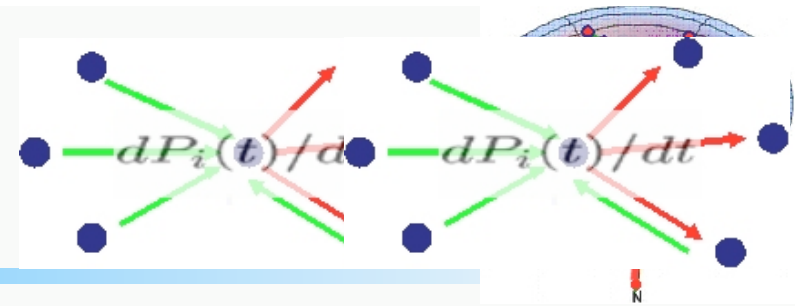
- Protein G and its mutants NuG1 and NuG2
  - Small, two-state folders
  - G was mutated to alter the hairpin formation order
  - Both have the same secondary and tertiary structure
- Our roadmaps captured the secondary structure formation order for Protein G and variants NuG1 and NuG2

[Thomas, Tang, Tapia, Amato JCB 07]

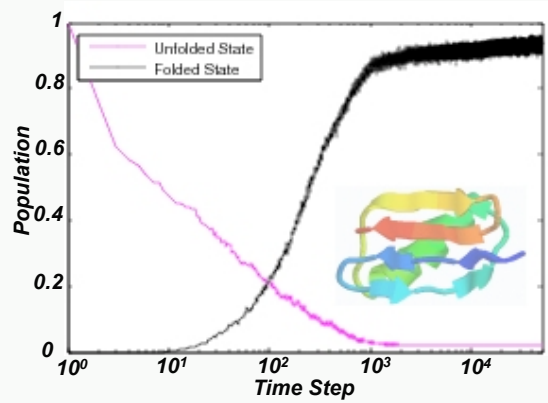
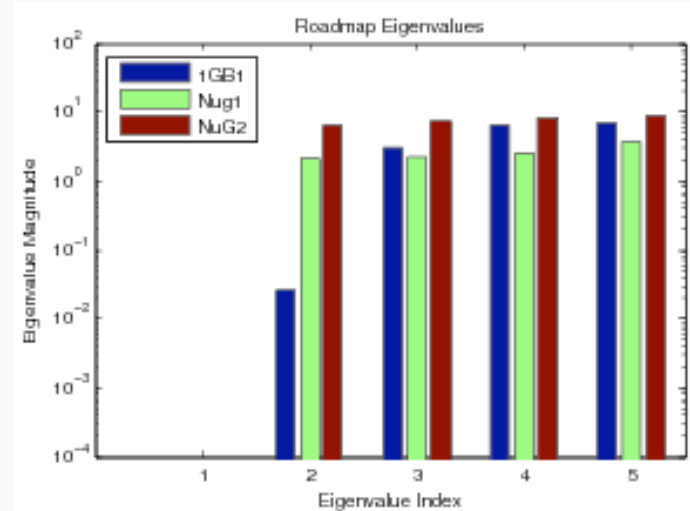


Mutants NuG1 and NuG2 fold 100 times  
faster than protein G [Nauli et al., 01]

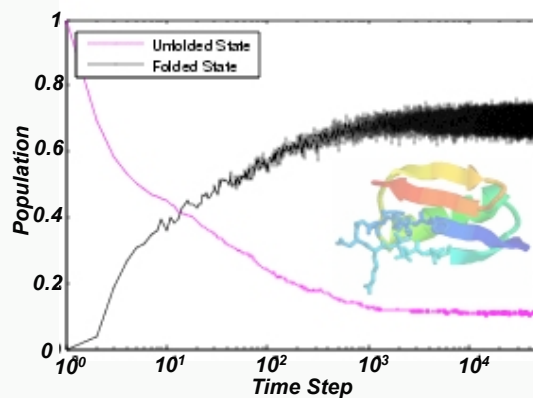
# Relative Rates of G, NuG1 and NuG2



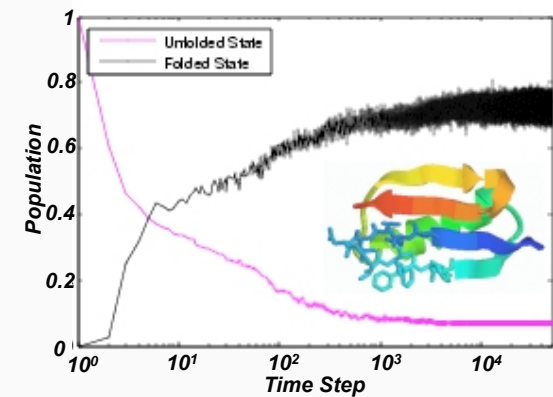
- MME: NuG1 and NuG2 faster than Protein G
- MMC: Faster folding rate of NuG1 and NuG2 also seen in population kinetics



**Protein G**



**NuG1**



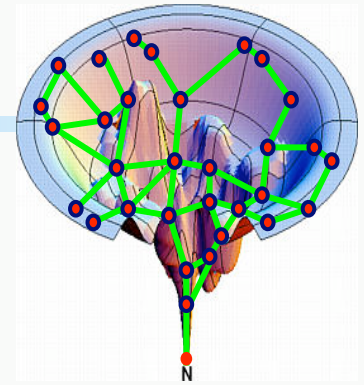
**NuG2**

# Summary

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## Map-based Protein Folding Techniques

Probabilistic Roadmap Methods for studying protein motions

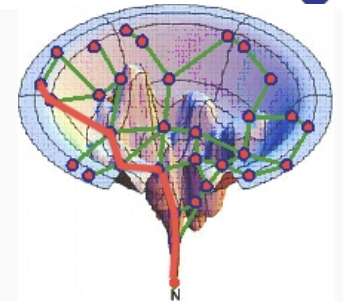
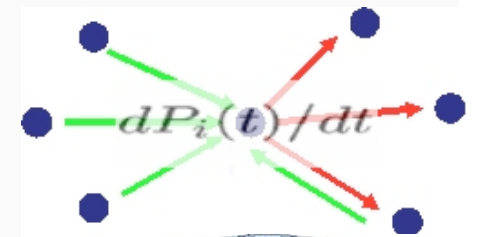


→ *Uses local transition probabilities to identify likely large-scale motions*

Technique 1: Map-Based Master Equation

Calculation (MME)

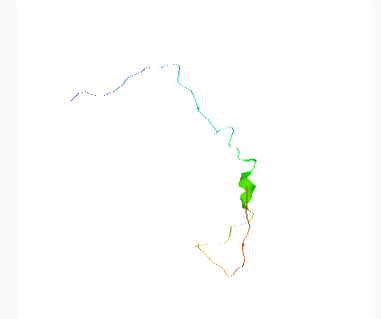
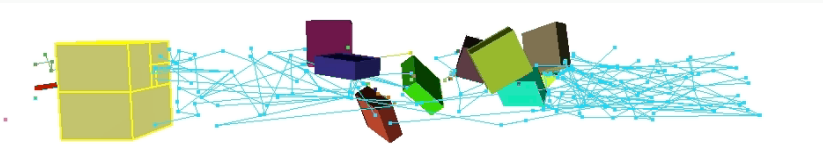
Technique 2: Map-Based Monte Carlo (MMC)



Ability to study time-based structural events

Ability to study a wide-range of structures and folding behaviors





# From Robots to Proteins...

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Many more results at: <http://parasol.tamu.edu/groups/amatogroup/foldingserver/>

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Luke Hunter, Protein Folding  
Kokil Jadika, Robotics  
Kasia Leyk, Protein Folding  
Lakshmi Reddy, Robotics  
Annette Stowasser, Protein Folding  
Manasi Vartak, Protein Folding

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Bryan Boyd, Texas A&M  
Prof. Marco Morales, ITAM  
Roger Pearce, Texas A&M  
Sam Rodriguez, Texas A&M  
Xinyu Tang, Google  
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